Stochastic changes affect Solanum wild species following autopolyploidization

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Abstract

Polyploidy is very common within angiosperms, and several studies are in progress to ascertain the effects of early polyploidization at the molecular, physiological, and phenotypic level. Extensive studies are available only in synthetic allopolyploids. By contrast, less is known about the consequences of autopolyploidization. The current study aimed to assess the occurrence and extent of genetic, epigenetic, and anatomical changes occurring after oryzaline-induced polyploidization of Solanum commersonii Dunal and Solanum bulbocastanum Dunal, two diploid (2n=2x=24) potato species widely used in breeding programmes. Microsatellite analysis showed no polymorphisms between synthetic tetraploids and diploid progenitors. By contrast, analysis of DNA methylation levels indicated that subtle alterations at CG and CHG sites were present in tetraploids of both species. However, no change occurred concurrently in all tetraploids analysed with respect to their diploid parent, revealing a stochastic trend in the changes observed. The morpho-anatomical consequences of polyploidization were studied in leaf main veins and stomata. With only a few exceptions, analyses showed no clear superiority of tetraploids in terms of leaf thickness and area, vessel number, lumen size and vessel wall thickness, stomata pore length and width, guard cell width, and stomatal density compared with their diploid progenitors. These results are consistent with the hypothesis that there are no traits systematically associated with autopolyploidization.

Key words: chromosome doubling, methylation, microsatellite, stomata, vessels, potato.

Introduction

Polyploidy is widespread in higher plants and represents a major evolutionary force (Ramsey and Schemske, 2002). Genome sequencing has provided evidence that even a number of species traditionally considered diploid (e.g. corn, Arabidopsis, rice) have experienced genome duplication events during their evolution. Polyploidization entails a reorganization of the full genome to enable harmonious cohabitation in the same nucleus of genes of different origin (in the case of allopolyploids) or of duplicated genes from identical or very similar genomes (in the case of autopolyploids). The structural and functional modifications induced by polyploidization are probably the key to the success of polyploid plants in nature. Indeed, it may greatly affect the morphology, physiology, and secondary metabolism of newly formed offspring.
For a specific trait, the phenotype could be similar to a progenitor, intermediate between the progenitors, or new (Comai et al., 2000). Possible morpho-anatomical and physiological polyplody effects include, among others, changes in cell and organ size (Schranz and Osborn, 2004), leaf width:length ratio (Stanys et al., 2006), and transpiration and/or photosynthetic rate (Levin, 2002). Polyplody can also cause the appearance of traits with a significant impact on crop production, such as resistance to stresses and a shift from sexual to apomictic reproduction (Quarin et al., 2001). Phenotypic changes caused by polyploidization are expected to be the result of genetic modifications. The first study demonstrating whole-genome changes after chromosome doubling was carried out in Brassica spp. by Song et al. (1995), who reported nuclear modifications in newly synthesized polyploids on the basis of restriction fragment length polymorphism patterns. Since then, similar phenomena have been documented in a wide range of plant species (reviewed by Yang et al., 2011). Ploidy-induced structural genomic changes are strongly associated with epigenetic alterations. The latter refer to heritable changes in gene expression that occur without alteration of the DNA sequence, such as DNA (cytosine) methylation, histone methylation, acetylation, phosphorylation, ubiquitylation, and sumoylation (Lauria and Rossi, 2011). Epigenetic changes often occur during genome merger and doubling as demonstrated, for instance, in wheat and Arabidopsis allopolyploids (reviewed by Chen, 2007).

Extensive molecular and phenotypic studies are available in several synthetic allopolyploids, including the wheat group (Triticum–Aegilops) (Levy and Feldman, 2004), Arabidopsis thaliana (L.) Heynh. (Comai et al., 2000), Tragopogon miscellus G. B. Ownbey (Tate et al., 2006), and Nicotiana spp. (Lim et al., 2006). This type of polyploidy is generally subjected to conspicuous and rapid genetic and epigenetic changes during the first generations and offers a preferential model to study the effects of polyploidization on phenotype and gene expression. Alterations may be necessary to overcome incompatibility between divergent genomes and have been considered important for the so-called initial ‘revolutionary phase’ of polyploid formation (Levy and Feldman, 2004). As autopolyploids arise by doubling of similar homologous genomes, it is believed that substantial alterations might not be required after their formation (Comai, 2005; Parisod et al., 2010). According to most findings reported in the literature, changes are less dramatic than those induced by allopolyploidization. Stupar et al. (2007) reported no DNA polymorphism within a ploidy series of Solanum phureja Juz. & Bukasov. Similarly, Allario et al. (2011) showed that microsatellite profiles of diploid and tetraploid Citrus limonia Osbeck were identical. Transcriptome and proteome analysis have generally confirmed the presence of only subtle changes in cabbage (Albertin et al., 2005), potato (Stupar et al., 2007), maize (Riddle et al., 2010), and Rangpur lime (Allario et al., 2011). Studies in A. thaliana (L.) Heynh. have corroborated the findings that stable, non-stochastic, developmentally specific gene expression alterations occur following autopolyploidization (Yu et al., 2010). In the same species, microarray analysis did not allow the identification of a reliable group of polyploidy-regulated genes (Pignatta et al., 2010). A recent paper by Li et al. (2012) reported a small but stable increase in the expression of cell-cycle genes (ICK1, ICK2, and ICK5) in autotetraploid seedlings of A. thaliana.

Given the evolutionary significance and practical breeding value of autopolyploidy and the scant coverage in the literature, the objective of this research was to assess the occurrence and extent of genetic, epigenetic, and anatomical changes occurring after artificial somatic polyploidization of two diploid (2n=2x=24) potato species, Solanum commersonii Dunal and Solanum bulbocastanum Dunal. Both species possess important resistance traits (to low temperatures and late blight, respectively) that can be exploited for breeding the tetraploid (2n=4x=48) cultivated potato. However, they are sexually isolated from Solanum tuberosum. Doubling their chromosome complement is an important prerequisite to sexually overcome incompatibility barriers (Carputo and Barone, 2005). Therefore, these species represent useful plant materials to compare the effects of early polyploidization at phenotypic and DNA levels.

Materials and methods

Plant material

Synthetic autotetraploids of S. bulbocastanum Dunal and S. commersonii Dunal were generated as described previously (Caruso et al., 2011). Briefly, a clone of diploid S. commersonii (PI 243503), coded cmm1t, and a clone of diploid S. bulbocastanum (PI 275190), coded blb1c, were subjected to oryzaline treatment to produce the autotetraploid genotypes used in this study. Those deriving from cmm1t were coded cmm15, cmm23, cmm24, cmm27, and cmm30. Those produced from blb1c were coded blb10, blb22, blb25, and blb26. To collect material for molecular and morpho-anatomical analyses, plants were micropropogated on MS medium (Murashige and Skoog, 1962) with sucrose (30 g l⁻¹) and microorgan (9 g l⁻¹)3, adjusted to pH 5.8, and then transferred into pots in a temperature-controlled greenhouse.

Molecular analysis

Fully developed young leaves of diploid progenitors and synthetic tetraploids were collected from individual plants on the same date. From each genotype, DNA was isolated from a leaf pool of three different plants using a DNeasy Plant mini kit (Qiagen, Valencia, USA) following the manufacturer’s instructions. DNA quality and integrity were checked by gel electrophoresis and spectrophometric assay. Simple sequence repeat (SSR) analyses were carried out with 12 nuclear microsatellite (SSR) primer pairs chosen from three sources: five primer pairs (STM1053, STM0031, STM1104, STM1052, and STM1106) from Milbourne et al. (1998), three (STM5114, STM5127, and STG0001) from Ghislain et al. (2009), and four (STI004, STI012, STI30, and STI32) from Feingold et al. (2005). These were recommended by the Centro Internacional de la Papa (http://www.cipotato.org) based on quality criteria, genome coverage, and locus-specific information. PCR reactions were performed in a 20 µl volume containing 1× reaction buffer with 2.5 mM MgCl₂, 0.2 mM of each dNTP, 25 pM carboxyfluorescein (FAM)-labelled forward SSR primer, 15 pM reverse SSR primer, 1 U of GoTaq DNA polymerase (Promega, Madison, USA), and 40 ng of genomic DNA. PCR was carried out using the following cycling profiles: 4 min at 94°C, 31 cycles of 45 s at 94°C, 1 min at annealing temperature, and 1 min at 72°C, with a final extension step of 5 min at 72°C. PCR products were separated on an ABI PRISM®
3130 DNA Analyzer. Size calibration was performed with the molecular weight ladder GenScan™ 500 ROX™ standard (Applied Biosystems, Foster City, USA). SSR alleles were detected and scored using GeneScan® Analysis software (Applied Biosystems).

To detect methylation patterns at the 5'-CCGG-3' sites, the methylation-sensitive amplification polymorphism (MSAP) technique was employed. Two methylation-sensitive isoschizomere enzymes, HpaII and MspI, were used, and a double digestion was set up according to the protocol of Reyna-López et al. (1997) and adapted by Xiong et al. (1999). Fifteen selective amplifications were performed for S. commersonii diploid and tetraploid genotypes, using two Fam-labeled EcoRI primers (E-TCCA and E-TCAA) combined with different HpaII/MspI (HM) primers: E-TCCA/HM-AAC; E-TCCA/HM-AAG; E-TCCA/HM-AG; E-TCCA/HM-AC; E-TCCA/HM-ATT; E-TCCA/HM-ACC; E-TCCA/HM-AGC; E-TCCA/HM-AC; E-TCCA/HM-AAC; and E-TCCA/HM-AAG. Seven selective amplifications were used for S. bulbocastanum diploid and tetraploid genotypes combining E-TCCA with the following HM primers: HM-AAC; HM-AAG; HM-ACA; HM-ACC; HM-ATC; HM-AGC; and HM-AGG. The GeneScan™ 500 ROX™ standard was used as size standard. SSR and MSAP fragments were separated electrophoretically using a capillary sequencer (ABI PRISM® 3130 Genetic Analyzer). A genetic profiler (GeneMapper®, Applied Biosystems) was used to display the results and to export data in text format. The intensity of the peaks was not taken into account, and fragments with identical mobility were considered to be monomorphic. For each genotype, polymorphic MSAP alleles were recorded as present (1) or absent (0). Methylation levels (hypo- and hypermethylation) were evaluated based on the different digestion patterns of the two methylation-sensitive isoschizomers, MspI and HpaII. All methylation changes in the CCGG site of diploid progenitors versus their tetraploid derivatives (including the appearance of new epi-alleles and the disappearance of parental epi-alleles) were calculated. For both SSR and MSAP analyses, DNA samples from each genotype were analysed in triplicate to provide technical replicates.

**Morphological and anatomical analysis**

Analyses were performed on diploid cmm1t and its tetraploids cmm23, cmm24, and cmm30, and on diploid blb1c and its tetraploids blb10, blb22, blb25, and blb26. Fully expanded leaves were collected at the same phenological stage (before flowering) on the same date. Digital images of the leaves were analysed using AnalySIS 3.2 (Olympus, Hamburg, Germany), a software program devised to quantify morpho-anatomical features. For anatomical analysis, from each genotype, three leaves were selected and fixed in 40% formaldehyde:glacial acetic acid:50% ethanol: 5:5:90 by vol-

To estimate better the predominant event between methylation and demethylation, a parameter termed methylation

**Results**

**Microsatellite analysis**

In diploid S. commersonii and its tetraploids, we were able to detect a total of 15 alleles (Table 1). Marker STM1052 did not amplify scorable alleles and was therefore not taken into account. Four loci (STG0001, STI004, STI012, and STM1053) were heterozygous, while the others were homozy
gous. On average, 1.4 alleles per locus were detected. For S. bulbocastanum, we were able to detect a total of 18 alleles by analysing 11 microsatellite loci. Marker STM0031 was ruled out from the analysis as it did not amplify scorable alleles. Seven loci were heterozygous: STM1052, STM1053, STM1104, STM1106, STG0001, STI004, and STI032. On average, we identified 1.4 alleles per locus. In neither species did the tested loci show polymorphisms between the diploids and the diploid progenitor they were derived from (data not shown), suggesting a lack of structural changes following polyploidization.

**MSAP analysis**

As reported in Table 2, due to the different sensitivities exhibited by the isoschizomers (McClelland et al., 1994), methyla
tion of the internal cytosine (C5mCGG, or CG sites) or the hemi- (single-strand) methylation of the external cytosine

(5mCGG, or CHG sites) was unequivocally distinguished (for details, see Supplementary Fig. S1 at JXB online). In particular, from the comparison of EcoRI/HpaII and EcoRI/MspI restrictions, four types of alteration in cyto
sine methylation at the CCGG sites of random genomic loci were identified: CG hypermethylation (hereafter coded as CG hyper), CG hypomethylation (coded as CG hypo), CHG hypermethylation (coded as CHG hyper), and CHG hypomethylation (coded as CHG hypo) (see Supplementary Fig. S2 at JXB online). In agreement with Ochogavia et al. (2009), to rule out confounding polymorphic bands due to mutations or methylation events, only ‘genuine epigenetic polymorphic markers’ (called unambiguous) that appeared in one but not in both HpaII and MspI digestions were scored for a given genotype. Differences detected between the diploid and synthetic tetraploids appeared to be stochastic (Table 2). In S. commersonii, out of 2545 epi-alleles, 87% did not change. To estimate better the predominant event between methylation and demethylation, a parameter termed methylation
The methylation ratio (MR) was calculated as the ratio between the hyper- and hypomethylation frequencies according to the following formula: MR=(CG hyper+CHG hyper)/(CG hypo+CHG hypo).
hyper incidences (5.2 for both). On average, the CHG were the sites changing the most (7.4 vs 6.0% of CG sites), with the exception of cmm15. MSAP analysis of *S. bulbocastanum* genotypes revealed no change for 94% of loci examined (Table 2). Also, in this species, all differences detected between the diploid blb1c and its synthetic tetraploids were stochastic. In blb10 and blb22, hypermethylation events were higher than hypomethylations (MR=1.2 and 2.3, respectively), whereas in blb25 and blb26 the opposite was true (MR=0.7 for both). On average, CG sites changed more than CHG (4.2% vs 2.2%), although in blb25, hyper- and hypomethylation at CHG sites occurred at the same frequency (2.3%). For both species, no change occurred concurrently in all tetraploids analysed with respect to their diploid parent, revealing a stochastic trend in the changes observed.

**Leaf morpho-anatomical analyses**

The quantitative leaf anatomical traits are summarized in Table 3. Both diploids and tetraploids had leaves with the typical dorsoventral structure with no evident qualitative modifications induced by polyploidization. In neither species did LT change significantly following polyploidization. The only exception was cmm30, whose LT increased significantly compared with diploid cmm1t. For *S. commersonii*, cmm23 and cmm30 this was similar to that in diploid cmm1t. By contrast, cmm24 showed a significantly lower LA. The same parameter was more variable in tetraploids of *S. bulbocastanum*. Indeed, compared with the diploid parent blb1c, blb22 showed a larger LA, blb10 and blb25 a similar LA, and blb26 a lower LA. Diploid cmm1t generally performed differently from its synthetic tetraploids in terms of xylem characteristics. Indeed, VN, VLA, VLD, and VWT in diploid *S. commersonii* were 51, 44, 25, and 15% higher than the mean values of its tetraploids, respectively (Fig. 1a, b; Table 3). A different trend was observed in *S. bulbocastanum* for VN and VLA. Indeed, the main veins of tetraploids generally comprised a greater number of larger vessels than their diploid counterparts (Fig. 1c, d; Table 3). VLD and VWT of diploid *S. bulbocastanum* did not deviate much from the mean value of its tetraploids. SEM analysis data are shown in Figs 2 and 3. In both species, SL did not change significantly following polyploidization. By contrast, SD underwent changes following genome doubling. They were stochastic in *S. commersonii* tetraploids with respect to cmm1t, whereas in *S. bulbocastanum*, the SD changes were more consistent, with all tetraploids showing lower values compared with blb1c (Fig. 3a, b). Diploid cmm1t behaved differently from its synthetic tetraploids in terms of SW (Fig. 3c, d) but not in terms of CW. By contrast, diploid blb1c was similar to its tetraploids in terms of SW but not in terms of CW.

**Discussion**

Studies on allopolyploids have contributed considerably to gaining insights into polyploidy-related processes. However, such polyploids inherently change two parameters at once, and hybridization rather than polyploidization per se may give rise to genome modifications (Doyle et al., 2008). By contrast, autopolyploids have the advantage of restricting the changes to one parameter, namely chromosome number, ruling out whole-genome alterations primarily connected with incompatible genomes. In this study, to understand better the chromosome doubling effects on genome stability, we tested the behaviour of SSR loci after polyploidization. None of the studied SSR loci demonstrated changes following genome doubling in microsatellite length. Our data not only confirmed the doubled diploid nature of our tetraploid genotypes but also revealed that structural changes due to polyploidization did not affect the genome of the two species analysed. Similar findings have also been reported by Ozkan et al. (2006) in *A. thaliana*, Stupar et al. (2007) in *Solanum* spp., and Allario et al. (2007).

### Table 3. Results (mean ±standard error) on leaf lamina thickness (LT) and area (LA), number of vessels per main vein (VN), vessel lumen area (VLA), and diameter (VLD), and vessel wall thickness (VWT) of *S. commersonii* and *S. bulbocastanum* synthetic tetraploids and the diploid parents they derived from (cmm1t and blb1c).

<table>
<thead>
<tr>
<th>Genotype</th>
<th>LT (µm)</th>
<th>LA (cm²)</th>
<th>VN (n)</th>
<th>VLA (µm²)</th>
<th>VLD (µm)</th>
<th>VWT (µm)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>S. commersonii</em></td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>cmm1t (2n=2x)</td>
<td>255.6±8.0 a</td>
<td>64.87±6.48 b</td>
<td>88.6±8.4 b</td>
<td>376.0±23.3 c</td>
<td>22.6±0.1 d</td>
<td>2.7±0.05 c</td>
</tr>
<tr>
<td>cmm23 (2n=4x)</td>
<td>245.1±4.7 a</td>
<td>59.47±6.37 b</td>
<td>45.5±5.20 a</td>
<td>267.7±16.0 b</td>
<td>19.9±0.0 c</td>
<td>2.7±0.06 c</td>
</tr>
<tr>
<td>cmm24 (2n=4x)</td>
<td>237.0±6.6 a</td>
<td>32.74±1.74 a</td>
<td>47.6±2.6 a</td>
<td>114.2±6.3 a</td>
<td>12.7±0.0 a</td>
<td>2.0±0.03 a</td>
</tr>
<tr>
<td>cmm30 (2n=4x)</td>
<td>281.8±8.5 b</td>
<td>56.47±6.88 b</td>
<td>39.0±4.2 a</td>
<td>246.1±18.5 b</td>
<td>18.3±0.0 b</td>
<td>2.2±0.05 b</td>
</tr>
<tr>
<td><em>S. bulbocastanum</em></td>
<td></td>
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<tr>
<td>blb1c (2n=2x)</td>
<td>243.0±7.9 ab</td>
<td>27.84±0.92 bc</td>
<td>33.3±2.3 a</td>
<td>390.9±24.1 a</td>
<td>23.5±0.1 a</td>
<td>3.1±0.07 b</td>
</tr>
<tr>
<td>blb10 (2n=4x)</td>
<td>222.8±9.2 ab</td>
<td>29.97±0.73 c</td>
<td>53.3±0.6 b</td>
<td>499.3±44.5 ab</td>
<td>27.1±0.1 a</td>
<td>2.7±0.06 a</td>
</tr>
<tr>
<td>blb22 (2n=4x)</td>
<td>256.1±10.1 b</td>
<td>34.00±1.51 d</td>
<td>52.0±1.5 b</td>
<td>483.0±47.0 ab</td>
<td>26.8±0.1 a</td>
<td>3.0±0.07 b</td>
</tr>
<tr>
<td>blb25 (2n=4x)</td>
<td>212.6±6.8 a</td>
<td>25.23±1.31 ab</td>
<td>46.0±1.2 b</td>
<td>445.0±27.8 ab</td>
<td>25.3±0.1 a</td>
<td>2.9±0.05 b</td>
</tr>
<tr>
<td>blb26 (2n=4x)</td>
<td>229.6±12.5 ab</td>
<td>22.81±0.64 a</td>
<td>36.0±5.6 a</td>
<td>512.0±42.2 b</td>
<td>26.4±0.0 a</td>
<td>3.1±0.08 b</td>
</tr>
</tbody>
</table>

a Each value of all measured parameters represents the mean of triplicate determinations.

b In each column, within each species, means denoted by the same letter did not differ significantly at P ≤0.05 according to Duncan's multiple range test.
et al. (2011) in Citrus ssp. Genome modifications have rarely been reported. A rapid elimination of DNA sequences was detected in induced autopolyploid of Phlox drummondii Hook by Raina et al. (1994). In a recent review, Parisod et al. (2010) suggested that, in synthetic autopolyploids, rapid elimination of DNA sequences may be possible, but this phenomenon is not so extensive during the first generations following genome doubling.

While genetic mutations can explain the cause of variability mainly over evolutionary time, epigenetic changes, which are potentially reversible, offer a flexible expedient for a polyploid cell to respond to genomic shock, especially in the early stages of polyploid formation, leading to various genetic variations. Alterations in cytosine methylation level were observed in our S. bulbocastanum and S. commersonii tetraploids versus their diploid parents (Table 2). In particular, various types of change were identified, including both hypo- and hypermethylation at CG and CHG sites, known to be the most affected targets of DNA methyltransferase (Law and Jacobsen, 2010). On average, the trend we observed was a subtle increase in methylation, although in all genotypes we detected both phenomena. Our data suggest that the S. commersonii and S. bulbocastanum epigenome landscape responds to autopolyploidization, probably to handle the consequences deriving from ploidy alterations. Ploidy-driven DNA methylation changes are likely to be more extensive than those we considered. Indeed, the real number of random loci undergoing methylation changes is probably underestimated, because, as suggested by Ochogavia et al. (2009), only a subset of MSAP markers (those called ‘genuine epigenetic polymorphic markers’) were prudentially analysed. Although our first-generation polyploids represent the plant materials to use in potato breeding, the DNA methylation changes occurring at the observed genetic loci have the potential to be inherited. Indeed, several studies suggest that epigenetic alterations generally persist through meiosis, showing a stable inheritance over many generations (reviewed by Hauser et al., 2011). In autotetraploid A. thaliana, Mittelsten Scheid et al. (2003) reported that epialleles lead to heritable gene silencing persisting after segregation from the inactivating allele. Investigating the mechanisms beyond polyploidy-associated transcriptional gene silencing in A. thaliana, Baubec et al. (2010) found that two epigenetic marks, namely DNA methylation and histone methylation, cooperate to give rise to a ‘double lock’ on transcriptional silencing, thus generating an extremely stable epigenetic state. For this reason, epialleles in polyploid plants might be highly relevant for long-term adaptation of gene expression patterns (Baubec et al., 2010). In our polyploids, the Alexander’s stain revealed a high pollen viability (>70% in all tetraploids; not shown), suggesting that it may be possible to obtain subsequent generations to study the effects of meiosis on the stability of epialleles.

Fig. 1. Light microscopy views of cross-sections of main veins in leaves of the diploid (a) and tetraploid (b) S. commersonii, and diploid (c) and tetraploid (d) S. bulbocastanum. Bar, 100 μm.
For both species, no change occurred concurrently in all tetraploids analysed with respect to their diploid parent, revealing a stochastic trend in the changes observed. The occurrence of stochastic methylation alterations in our genetically identical autopolyploids confirms results previously reported in maize (Zhao et al., 2007). Such a phenomenon demands additional studies on subsequent generations to verify whether the identified methylation variations are artefacts of the lack of meiosis. Ploidy-induced DNA methylation changes were reported by Ochogavia et al. (2009) in Eragrostis curvula Schrad. Nees and Lavania et al. (2012) in Cymbopogon spp. The authors reported enhanced cytosine methylation as a result of genome duplication. By contrast, in Paspalum notatum Flügge (Martelotto et al., 2007) and watermelon (Wang et al., 2009), DNA methylation sites were similar between diploids and autotetraploids. These reports, along with our results, suggest that epigenetic modifications are probably induced by genome doubling per se but appear to be linked to different taxa and might be related to (i) the plastic nature of the epigenetic landscape of the species, which tends to respond more or less promptly to external and internal stimuli; and (ii) the CG/CHG content of a given species, as those rich in either CG or CHG may depend on methylation more heavily than species with lower CG/CHG content to regulate gene expression following polyploidization.

Despite their importance, the morphological and anatomical effects of alterations in ploidy level are not well understood (Riddle et al., 2006), and how ploidy and cell size are linked is still unclear (Tsukaya, 2008). In general, polyploidy results in significant cell enlargement, probably

**Fig. 2.** Results of stomata pore length (SL, μm), stomata density (SD, pores mm⁻²), stomata pore width (SW, μm), guard cell width (CW, μm) of *S. commersonii* and *S. bulbocastanum* synthetic tetraploids and the diploid parents they were derived from (cmm1t and blb1c). Each value represents the average number of ten determinations (mean ± standard error). Means denoted by the same letter did not differ significantly at *P* ≤ 0.05 according to Duncan’s multiple range test.
to maintain a constant balance between the nuclear and cytoplasm volume (Cavaleri and Smith, 1985). Phenotypic analysis at the cellular level has demonstrated that synthetic polyploidy resulted in larger epidermal and guard cells in Cymbopogon spp. (Lavania et al., 2012) and Nicotiana spp. (Anssour et al., 2009), larger mesophyll cells in P. drummon- dii Hook (Vyas et al., 2007), and thicker leaves and larger mesophyll cells in Citrus spp. (Romero-Aranda et al., 1997). Polyploidization has also been considered responsible for changes at the vascular level in Manihot esculenta Crantz, including the formation of smaller xylem elements characterized by thinner cell walls (Nassar et al., 2008). In our research, we focused on characterizing the main veins and stomatal parameters in leaves, as they are related to water conductivity, photosynthetic capacity, and hydraulic vulnerability under drought conditions (Blackman et al., 2010). In general, our analyses did not show a consistent superiority of tetraploids compared with their diploid progenitors (Table 3, Figs 1a, b, 2, 3c, d). Exceptions were observed in S. commersonii for leaf LT and in S. bulbocastanum for VN and VLA (Fig. 1c, d) and CW (Fig. 1a, b). Lack of consistent phenotypic superiority of synthetic tetraploids on their diploid parents has been reported for a number of anatomic traits in potato (Stupar et al., 2007), Citrus limonia Osbeck (Allario et al., 2011), Artemisia annua L. (Banyai et al., 2010), and Atriplex confertifolia Torr. et Frém (Warner and Edwards, 1989). We hypothesize that the occasional tetraploid superiority we found may be due to genetic variation on the phenotypic effects of autopolyploidization. To explain the lack of phenotypic superiority of several tetraploids over their diploid parents, at least two hypotheses can be formulated. Firstly, plants with a higher ploidy may experience fitness costs due to elevated DNA content and replication, defective mitosis, and increased cell size (Stupar et al., 2007). To support this hypothesis further, a recent treatment by Tsukaya (2008) reported that an increase in ploidy might alter the ratio between components of the cell with different dimensions, adding stress to polyploid cell activity. Secondly, the epigenetic changes detected in our synthetic polyploids, albeit subtle, may have caused modifications in the expression of relevant genes. The chance that small methylation changes are likely to exert an effect on the phenotype has been described previously (Gemma et al., 2009; Einstein et al., 2010). Therefore, it would be interesting to study the whole-genome expression patterns of our diploid and tetraploid materials as well as other mechanisms regulating gene expression, such as histone modifications and chromatin states.

Fig. 3. Scanning electron microscopy results showing 2× versus 4× differences in terms of stomata pore width (a, b) and density (c, d) in S. commersonii (cmm) and S. bulbocastanum (blb), respectively.
In conclusion, our results on genome reorganization following chromosome doubling indicated that structural changes did not occur in the wild potato genotypes analysed here. By contrast, methylation modifications were found, with possible effects on gene silencing or activation that deserve further investigation. In terms of leaf anatomical characteristics, polyploidization did not result in a consistent and clear phenotypic superiority of tetraploids. Although large tetraploid/diploid combinations should confirm these data, they support the idea that studies on autoploidy should assume the null hypothesis that there are not traits systematically associated with autoploidy and, based on this assumption, that genome multiplication per se may be an evolutionary advantage (Parisod et al., 2010).

**Supplementary data**

Supplementary data are available at JXB online.

**Supplementary Fig. S1.** Possible methylation status at 5'-CCGG sites, and the corresponding banding patterns produced by the isoschizomers HpaII and MspI.

**Supplementary Fig. S2.** Examples of MSAP electropherograms and banding patterns on 2× and 4× genotypes of *S. commersonii*.

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**References**


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